#### Spatial patterns in a population model structured by cell size, quiescence and sensing radius

#### Hideki Murakawa

(Kyushu University, Japan)

joint work with

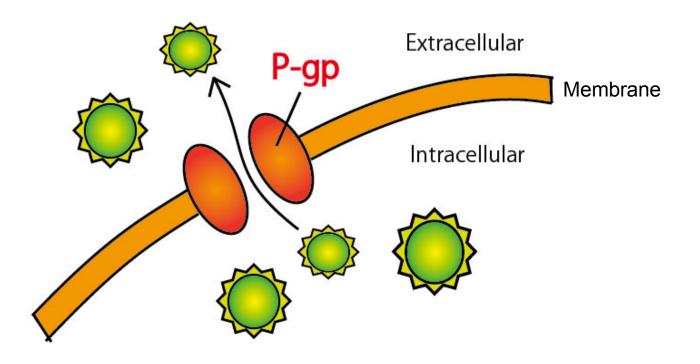
Arnaud Ducrot (Universite Victor Segalen Bordeaux 2, France)
Frank Le Foll (University of Le Havre, France)
Pierre Magal (Universite Victor Segalen Bordeaux 2, France)
Jennifer Pasquier (University of Le Havre, France)
Glenn F. Webb (Vanderbilt University, USA)

#### EVERYTHING DISPERSES TO MIAMI

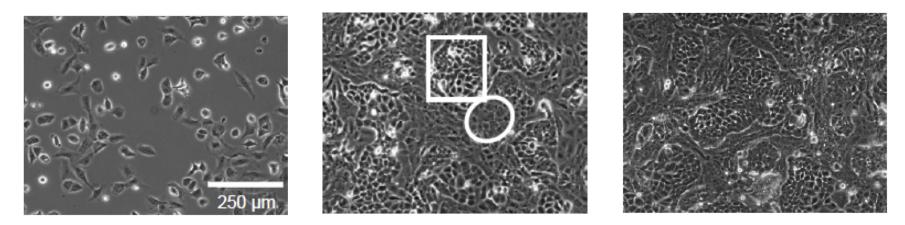
THE ROLE OF MOVEMENT AND DISPERSAL IN SPATIAL ECOLOGY, EPIDEMIOLOGY AND ENVIRONMENTAL SCIENCE December 14, 2012 The University of Miami Coral Gables, Florida

### Multidrug resistance, P-glycoprotein

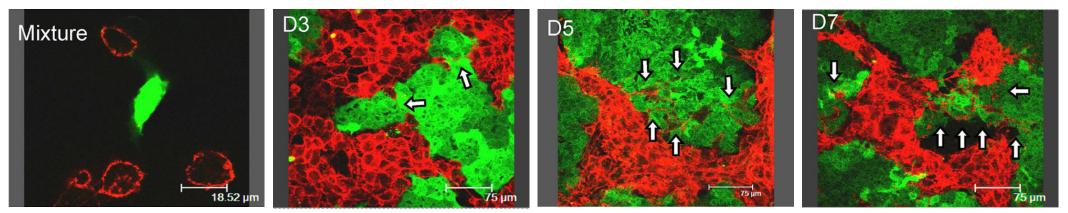
Multi-drug resistance is a phenomenon by which tumor cells exhibit resistance to a variety of chemically unrelated chemotherapeutic drugs. The classical form of multidrug resistance is connected to overexpression of membrane P-glycoprotein (P-gp), which acts as an energy dependent drug efflux pump.



### **Spatial behavior and P-gp transfers**



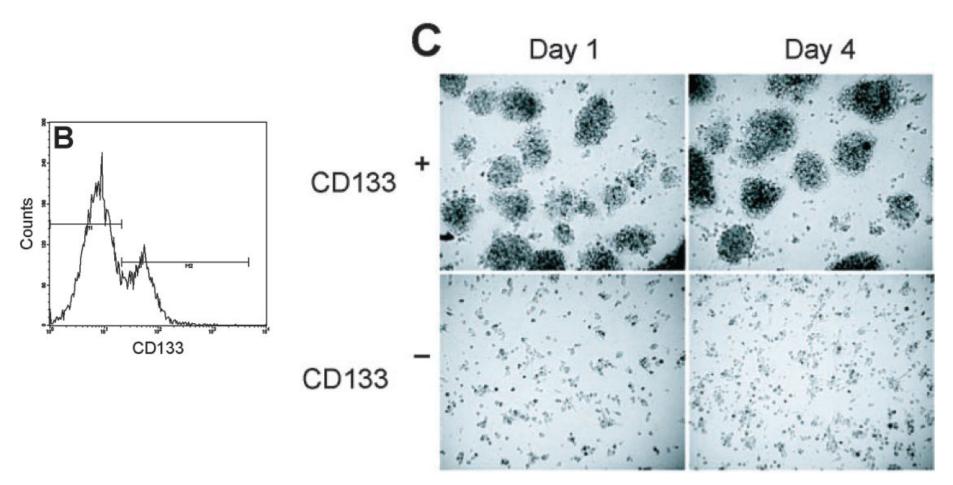
To obtain phase contrast micrographs of growing MCF-7 variants in co-cultures, dishes were seeded with a 50:50 mixture of MCF-7:MCF-7/DOXO at day 0. Morphological differences permit an immediate identification of each cell subpopulation. MCF-7 appeared birefringent and round (boxes) whereas MCF-7/DOXO are more flat and spread (ellipses). Note that the cells remained organized in well-delimited islets. Pasquier, Magal, Boulangé-Lecomte†, Webb & Le Foll ('11).



**Direct immunodetection of P-gp transfers in co-cultures of sensitive (MCF-7) and resistant (MCF-7/Doxo) variants of the human breast cancer cell line.** Mixtures of 50:50 ctgMCF-7:MCF-7/Doxo were co-cultured on glass coverslips during periods varying from 0 to 7 days (D0-D7). P-gp was immunodetected with phycoerythrin-conjugated (*PE*)-UIC2 mAb (*red fluorescence*) by confocal laser scanning microscopy in non-dispersed. From D3 to D7, sensitive ctgMCF-7 show an increasing P-gp-specific red membrane staining (*arrows*), restricted to the plasma membrane, in non-dispersed as well as in dissociated cells.

Pasquier, Galas, Boulangé-Lecomte, Rioult, Bultelle, Magal, Webb and Le Foll ('12).

### **Pattern & Proliferation**



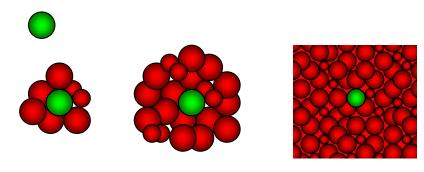
**B**, flow cytometry histogram in representative medulloblastoma tumor cells, with the first peak (gate M1) representing cells negative for CD133- phycoerythrin expression, and the second peak (gate M2) representing CD133 positive cells. Tumor cells were then sorted for CD133 expression by magnetic bead cell sorting. CD133+ and CD133- populations were collected, checked for purity by flow cytometry, and cultured separately in TSM for stem cell assays. Purity was found to range from 46.9 to 79.8% in CD133+ populations, and 92.6 to 97.3% in CD133- populations.

*C*, CD133+ tumor cells proliferated in culture as nonadherent spheres, whereas CD133- tumor cells adhered to culture dishes, did not proliferate and did not form spheres.

Singh, Clarke, Terasaki, Bonn, Hawkins, Squire, & Dirks ('03).

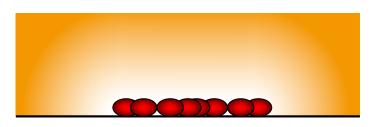
# **Pressure for growth** (Nonlocal pressure) $\overline{p}(t,x) = \int_{\Omega} K(x,y)p(t,y)dy.$

•Contact inhibition of growth

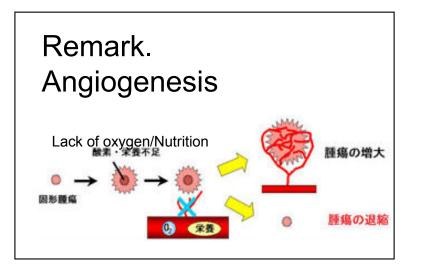


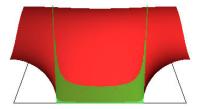
Sufficient supply

•Supply and demand for Nutrition or Oxygen

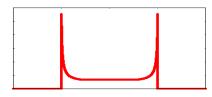


Excess of demand over supply





Numerical simulation for Nutrition-Absorption relation



A function given by the nonlocal reaction (defined below)

#### A model incorporating Cell size, Fission, Quiescence & Sensing radius.

$$\begin{split} u(t,x,s) &: \text{population density of proliferating cells.} & t: \text{time,} \\ v(t,x,s) &: \text{population density of quiescent cells.} & s: \text{spatial position,} \\ m(t,x) &= \int_{0}^{+\infty} s\left[u+v\right](t,x,s)ds &: \text{density of mass.} \\ p(t,x) &= m(t,x) : \text{ pressure for motility,} \quad \overline{p}(t,x) = \int_{\Omega} K(x,y)m(t,y)dy: \text{ pressure for growth.} \\ & \begin{cases} u_t = \underbrace{\operatorname{div}_x \left(u \nabla_x p\right)}_{\operatorname{cell motility}} - \underbrace{\partial_s \left[g(s)u\right]}_{\operatorname{cell size growth}} & - \underbrace{\beta_u \left(\overline{p}(t,x),s\right)u}_{\operatorname{to quiescence}} + \underbrace{\beta_v \left(\overline{p}(t,x),s\right)v}_{\operatorname{from quiescence}} \\ & + \underbrace{4b\left(2s\right)u(t,x,2s\right) - b(s)u(t,x,s)}_{\operatorname{fission}} - \underbrace{\mu(s)u(t,x,s)}_{\operatorname{mortality}} \\ v_t = \underbrace{\operatorname{div}_x \left(v \nabla_x p\right)}_{\operatorname{cell motility}} + \underbrace{\beta_u \left(\overline{p}(t,x),s\right)u}_{\operatorname{from proliferation}} - \underbrace{\beta_v \left(\overline{p}(t,x),s\right)v}_{\operatorname{mortality}} \\ & t > 0, x \in \Omega, s > 0. \end{cases} \\ g(s) : \text{ cell size growth rate,} & \mu(s) : \text{ mortality rate,} \end{split}$$

 $\beta_u(p, s) : \text{ non-decreasing function for transition form proliferation to quiescence,} \\ \beta_v(p, s) : \text{ non-increasing function for transition form quiescence to proliferation,} \\ b(s) : \text{ division rate s.t. } b(s) = \begin{cases} 0 & \text{if } s \leq s_f, \\ \geq 0 & \text{if } s_f < s. \end{cases} \\ \text{ The cells cannot divide before they have reached a size } s_f \text{ .} \end{cases}$ 

### Fast transition

We assume that the dynamics of transition to and from quiescence and proliferation are fast compared to the other dynamics.

$$\begin{cases} u_t = \operatorname{div}_x \left( u \nabla_x p \right) - \partial_s \left[ g(s) u \right] - \varepsilon^{-1} \beta_u \left( \overline{p}(t, x), s \right) u + \varepsilon^{-1} \beta_v \left( \overline{p}(t, x), s \right) v \\ + 4b \left( 2s \right) u(t, x, 2s) - \left( b(s) + \mu(s) \right) u(t, x, s) \\ v_t = \operatorname{div}_x \left( v \nabla_x p \right) + \varepsilon^{-1} \beta_u \left( \overline{p}(t, x), s \right) u - \varepsilon^{-1} \beta_v \left( \overline{p}(t, x), s \right) v - \mu(s) v(t, x, s), \\ & \text{where } 0 < \varepsilon << 1. \end{cases}$$

Taking a formal limit as  $\varepsilon \searrow 0$ ,  $\beta_u (\overline{p}(t,x),s) u = \beta_v (\overline{p}(t,x),s) v$ .

$$u(t, x, s) = G\left(\overline{p}(t, x), s\right) n(t, x, s).$$

Set n(t, x, s) := (u + v) (t, x, s), : population density of total cells of size s.  $G(\overline{p}(t, x), s) := \frac{\beta_v (\overline{p}(t, x), s)}{\beta_u (\overline{p}(t, x), s) + \beta_v (\overline{p}(t, x), s)}.$ 

$$n_t = \operatorname{div}_x (n\nabla_x p) - \partial_s \left[ g(s)G\left(\overline{p}(t,x),s\right) n(t,x,s) \right] +4b \left(2s\right) G\left(\overline{p}(t,x),2s\right) n(t,x,2s) - b(s)G\left(\overline{p}(t,x),s\right) n(t,x,s) -\mu(s)n(t,x,s).$$

Equation for density of mass  

$$n_t = \operatorname{div}_x (n \nabla_x p) - \partial_s [g(s)G(\overline{p}(t,x),s) n(t,x,s)]$$
  
 $+4b(2s) G(\overline{p}(t,x),2s) n(t,x,2s) - b(s)G(\overline{p}(t,x),s) n(t,x,s)$   
 $-\mu(s)n(t,x,s).$ 

We assume that

$$g(s) = gs, \quad G(\cdot, s) = G(\cdot), \quad \mu(s) = \mu.$$

Multiply both sides by s and integrate over  $(0, +\infty)$  w.r.t. s to obtain Note  $m(t, x) = \int_0^{+\infty} s [u + v] (t, x, s) ds$  $m_t = \operatorname{div}(m\nabla m) + G(\overline{m}) \int_0^{\infty} A(\cdot, \cdot, s) ds - \mu m.$ 

Here,

$$A(t, x, s) := -s\partial_s \left[ g(s)n(t, x, s) \right] + 4sb(2s)n(t, x, 2s) - sb(s)n(t, x, s).$$

 $\begin{array}{l} \mbox{Equation for density of mass}\\ A(t,x,s) := -s\partial_s \left[g(s)n(t,x,s)\right] + 4sb\left(2s\right)n(t,x,2s) - sb(s)n(t,x,s).\\ \mbox{Note} \quad b\left(s\right) = \left\{ \begin{array}{ll} 0 & \mbox{if } s \leq s_f,\\ \geq 0 & \mbox{if } s_f < s. \end{array} \right. \end{array}$ 

Therefore, 
$$\int_0^\infty A(s)ds = gm.$$

#### Simplified Model

$$m_t = \operatorname{div}(m\nabla m) + \left(gG\left(\int_{\Omega} K(\cdot, y)m(t, y)dy\right) - \mu\right)m.$$

### **Assumptions**

(P) 
$$\begin{cases} m_t - \Delta \phi(m) = F(\overline{m}) m & \text{in } (0, \infty) \times \Omega, \\ \frac{\partial \phi(m)}{\partial \nu} = 0 & \text{on } (0, \infty) \times \partial \Omega, \\ m(0, \cdot) = m_0 & \text{in } \Omega, \end{cases}$$

where  $\phi(s) = \frac{1}{2}s^2$ ,  $F(s) = gG(s) - \mu$ ,  $g, \mu > 0$ : const.  $\overline{m}(t, x) = \int_{\Omega} K(x, y)m(t, y)dy.$ 

•  $m_0 \in L^1_+(\Omega)$ ,

•  $K \in L^{\infty}_{+}(\Omega \times \Omega),$ 

•  $G: [0,\infty) \to [0,1]$  is Lipschitz continuous.

### Definition

**Definition.** (Weak energy solution) A measurable function  $m : [0, \infty) \times \Omega \to \mathbb{R}_+$  is said to be a weak energy solution of (P) if for each T > 0

(i) 
$$m \in L^2(Q_T)$$
 and  $w = \phi(m) \in L^2(0, T; H^1(\Omega))$ ,

(ii) *m* satisfies for each  $\eta \in C^1(\overline{Q_T})$  s.t.  $\eta(T, \cdot) \equiv 0$ 

$$\int_{Q_T} \left( \nabla w \cdot \nabla \eta - m\eta_t \right) dt dx = \int_{\Omega} m_0(x) \eta(0, x) dx + \int_{Q_T} \eta F\left(\overline{m}\right) m dt dx.$$

### **Existence and Uniqueness**

#### Theorem (Ducrot-Le Foll-Magal-M-Pasquier-Webb ('11))

For each  $m_0 \in L^3_+(\Omega)$  there exists a unique energy solution  $m \equiv m(t, x; m_0)$  of (P) such that

 $m \in L^{\infty}_{\text{loc}}\left([0,\infty); L^{3}(\Omega)\right) \cap C\left([0,\infty); L^{1}(\Omega)\right).$ 

Moreover for each M > 0 and each T > 0 there exists  $\delta = \delta(T, M) > 0$  such that for each  $m_0, m_1 \in L^3_+(\Omega)$ , if  $\|m_0\|_{L^1} \leq M$  and  $\|m_1\|_{L^1} \leq M$ , then

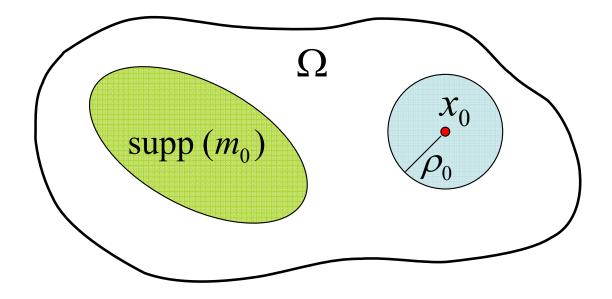
 $\|m(t,.;m_0) - m(t,.;m_1)\|_{L^1} \le \delta(T,M) \|m_0 - m_1\|_{L^1}, \quad \forall t \in [0,T].$ 

### Finite speed of propagation

#### Theorem (Ducrot-Le Foll-Magal-M-Pasquier-Webb ('11))

If  $m_0 \in L^3_+(\Omega)$  satisfies that there exists  $x_0 \in \Omega, \ \rho_0 \in (0, \text{dist } (x_0, \partial \Omega), \ m_0(x) = 0, \ a.e. \ x \in B(x_0, \rho_0),$ then there exists  $T^* > 0$  and a mapping  $\rho : [0, T^*] \to [0, \rho_0]$  such that  $m = m(t, x; m_0)$  satisfies

$$m(t,x) = 0$$
 for  $t \in [0,T^*], x \in B(x_0,\rho(t)).$ 



### **Numerical experiments**

$$m_t = \frac{1}{2}\Delta m^2 + \left(gG\left(\int_{\Omega} K_r(\cdot, y)m(t, y)dy\right) - \mu\right)m.$$

with periodic b.c.

β<sub>u</sub>(p) β<sub>v</sub>(p)

0.3

0.4

0.2

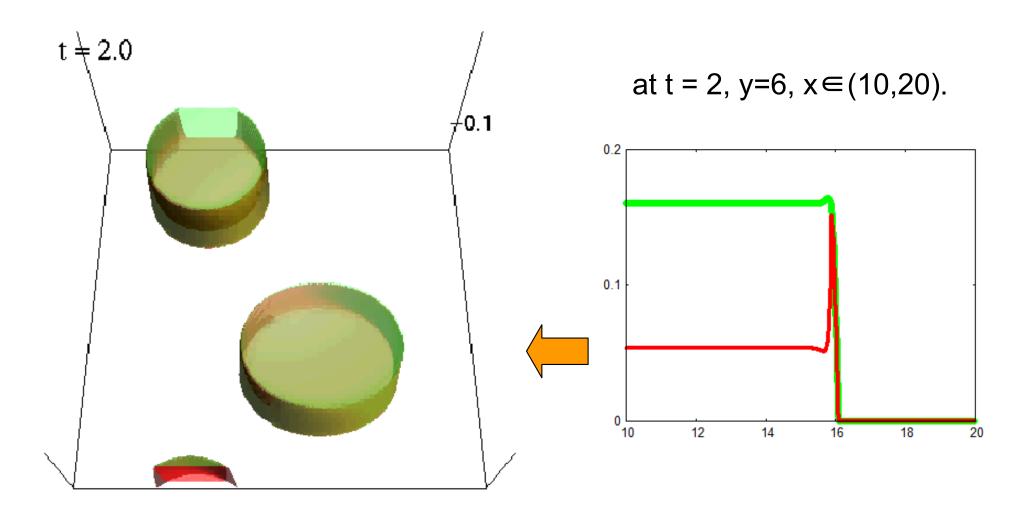
р

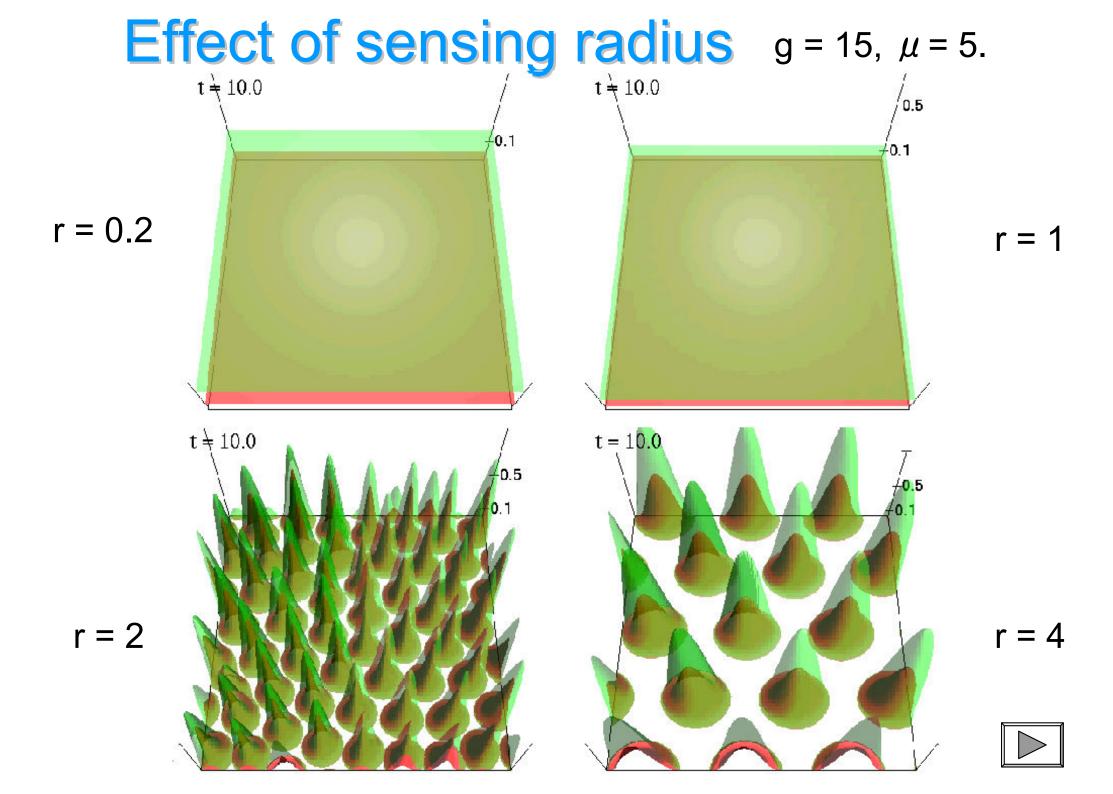
$$\Omega = (0, 20)^{2}.$$

$$G(p) := \frac{\beta_{v}(p)}{\beta_{u}(p) + \beta_{v}(p)}.$$

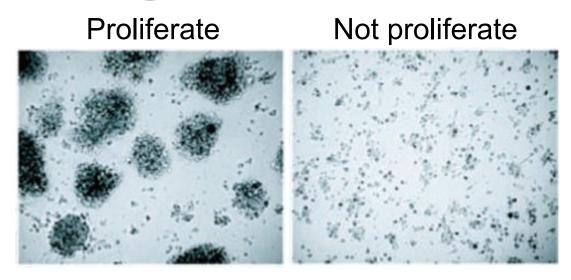
$$K_{r}(x, y) = \begin{cases} 1/(\pi r^{2}) & \text{if } |x - y| < r, \\ 0 & \text{otherwise.} \end{cases}$$

### Numerical experiments $g=15, \mu=5, r=0.2.$ m m m = m

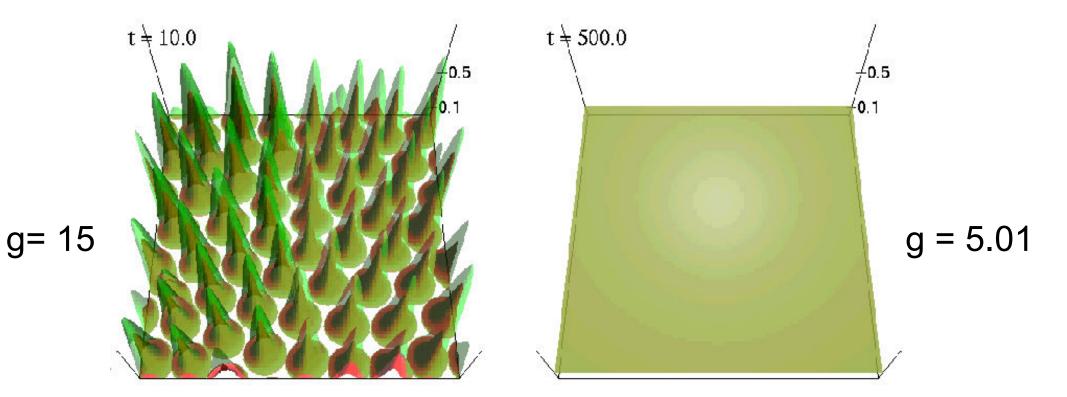


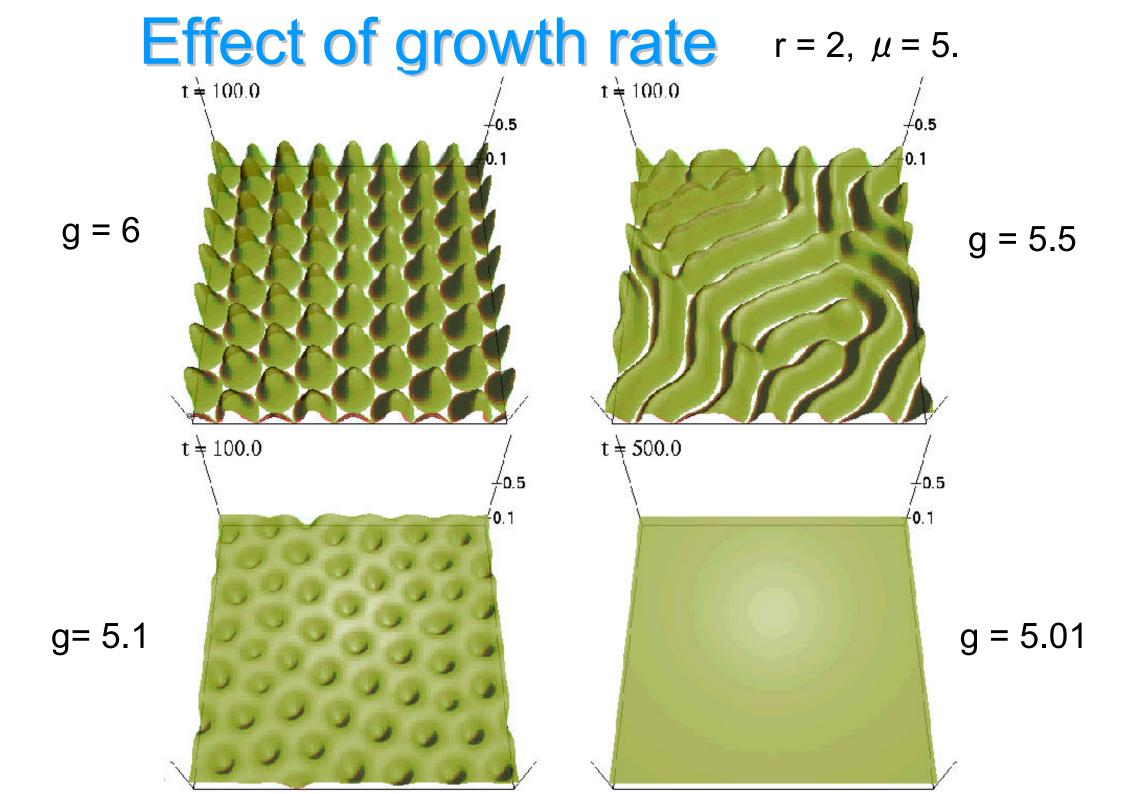


### **Effect of growth rate** $r = 2, \mu = 5.$

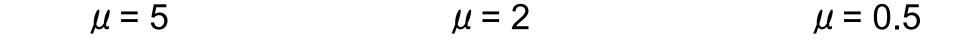


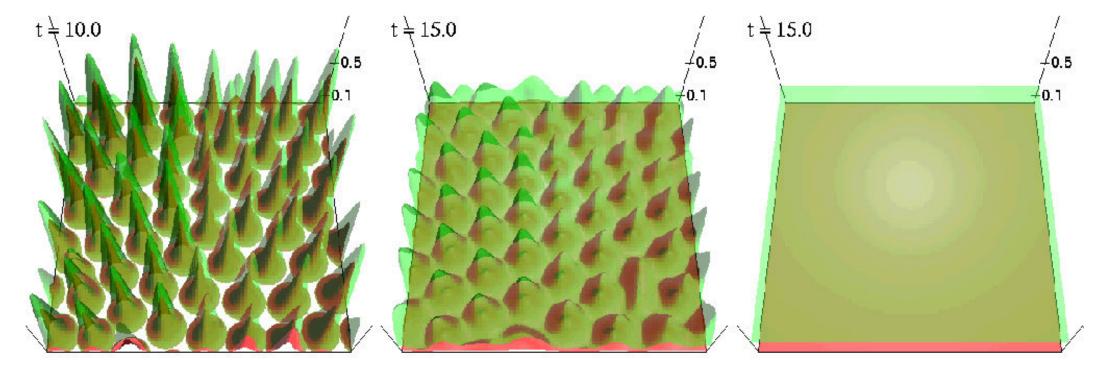
Singh, Clarke, Terasaki, Bonn, Hawkins, Squire, & Dirks ('03).





#### **Effect of mortality rate** r = 2, g = 15.

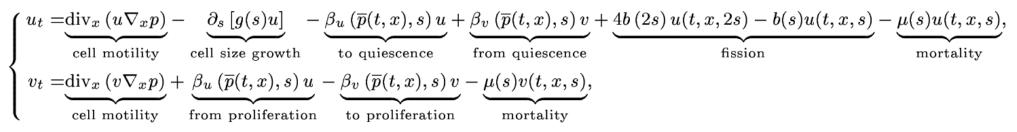




## Conclusion

#### A model incorporating Cell size, Fission, Quiescence & Sensing radius.

Contact inhibition,
 Supply and demand for Nutrition or Oxygen.



# Simplified Model $m_t = \frac{1}{2}\Delta m^2 + \left(gG\left(\int_{\Omega} K_r(\cdot, y)m(t, y)dy\right) - \mu\right)m.$

#### **Numerical results**

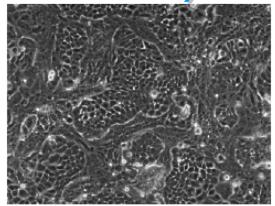
#### Theoretical results

Unique existence of the solution.Finite speed of propergation.

	Large	Small
Sensing radius	colonies	uniform distribution
growth rate		
mortality rate		

## Future work (Co-cultured cells)

$$\begin{cases} \frac{\partial m_1}{\partial t} = & \operatorname{div} \left( m_1 \nabla (a_{11} m_1 + a_{12} m_2) \right) \\ & + \left[ g_1 G_1 \left( \int_{\Omega} K_{r_1}(\cdot, y) (b_{11} m_1(\cdot, y) + b_{12} m_2(\cdot, y)) dy \right) - \mu_1 \right] m_1, \\ \frac{\partial m_2}{\partial t} = & \operatorname{div} \left( m_2 \nabla (a_{21} m_1 + a_{22} m_2) \right) \\ & + \left[ g_2 G_2 \left( \int_{\Omega} K_{r_2}(\cdot, y) (b_{21} m_1(\cdot, y) + b_{22} m_2(\cdot, y)) dy \right) - \mu_2 \right] m_2. \end{cases}$$



Pasquier, Magal, Boulangé-Lecomte†, Webb & Le Foll ('11).

